

Original Research Article

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Integrated Management of Major Foliar and Soil-Borne Diseases of Peanut (*Arachis hypogaea* L.) with Fungicides, *Trichoderma* and Castor Cake

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ABSTRACT

A field experiment was conducted at ICAR-DGR, Junagadh in *kharif* (rainfed or monsoon season crop) of 2009 and 2010 for integrated disease management (IDM) of major peanut diseases. The IDM component comprised of a combination of seed treatment with *Trichoderma*, mancozeb, tebuconazole; soil application of *Trichoderma* and castor cake; and foliar sprays of hexaconazole and tebuconazole. Observations on foliar fungal diseases were recorded by adopting 1-9 modified scale and in the case of soil borne diseases per cent incidence was recorded. The treatments were found effective to maintain the initial plant population and were highest in seed treatment with mancozeb followed by seed treatment with *Trichoderma harzianum*. Lowest incidence of soil-borne diseases was recorded in seed treatment with mancozeb and seed treatment with tebuconazole in comparison to untreated control. The foliar disease severity was lowest in treatment tebuconazole and two foliar sprays of tebuconazole whereas it was recorded highest in the untreated control. Highest pod and haulm yield were recorded in seed treatment with mancozeb and two sprays of hexaconazole and seed treatment with tebuconazole and two foliar sprays of tebuconazole whereas it was lowest in the untreated control. The cost of treatment are very important to manage diseases, therefore, it is suggested that combination of treatments are most suitable for disease management.

Keywords

Arachis hypogaea,
Biocontrol,
Integrated disease
management,
Peanut diseases and
Triazole fungicide.

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Introduction

Peanut or groundnut (*Arachis hypogaea* L.) is an important crop with high levels of proteins, carbohydrates, vitamins and minerals contained within seeds (Moss and Rao, 1995). While the cultivation of peanut may occur over a wide range of climatic conditions, soil type, temperature, and amount of rainfall and its distribution during crop season need special attention. Well-drained, sandy soils are best suited for peanut production (Beasley

et al., 1997). In 2013, the world peanut production was 45.7 million tonne from 25.4 million ha area with an average productivity of 1796.2 kg ha⁻¹ (FAO, 2013).

In India, peanut is an important oilseed crop, contributing about 36 per cent of the total oilseed production. India is the largest grower and second largest producer of peanut in the world. It is cultivated as *Kharif* (rainfed or

monsoon season crop) and *Rabi*-summer (irrigated, sown in winter and harvested in the spring) crop and the major peanut growing states are Gujarat, Andhra Pradesh, Tamil Nadu, Karnataka, Rajasthan, and Maharashtra which contributes around 90% of area and production. It is cultivated in 5.5 million ha area, divided in *Kharif* (4.6) and *Rabi* (1.0) with a production of about 9.7 million tonne (*Kharif*- 7.8 and *Rabi*- 1.9) and productivity 1750 kg ha⁻¹. The average yield of *Rabi*-peanut is around 1929 kg ha⁻¹, whereas *Kharif*-peanut is around 1712kg ha⁻¹ which is lower than major peanut growing countries (GOI, 2014). This may be attributed to the rainfed- cultivation of this crop coupled with damage caused by several pests and diseases. Among foliar fungal diseases, only a few are economically important such as early and late leaf spots commonly called as '*tikka*' disease and rust. Early leaf spot (ELS) is caused by *Cercosporaarachidicola* Hori (Perfect stage-*Mycosphaerella arachidis* Deighton and late leaf spot (LLS) is caused by *Phaeoisariopsispersonata* (Berk & Curt) V. Arx (Perfect stage – *Mycosphaerella berkeleyi* Jenkins). Rust is caused by *Puccinia arachidis* Speg. Among seed and soil-borne diseases, collar rot also called crown rot or seedling blight (*Aspergillus niger* Van Tieghem, *A. pulverulentus* (McAlp) Thom), stem rotor *Sclerotium* wilt (*Sclerotium rolfsii* Sacc), dry wilt or dry root rot (*Macrophomina phaseolina* (Tassi) Goid=*Rhizoctonia bataticola* (Taub.) Butler) have been recognized as major diseases. Economically important virus diseases are peanutbud necrosis disease (PBNB), peanut stem necrosis disease (PSND), peanut mottle and peanut clump. Peanut bud necrosis is sap transmissible and vectored by *Thripspalmi* Karny. Peanut mottle is transmitted in a non-persistent manner by aphids, *Aphis craccivora* and *Myzus persicae*. In India, among foliar fungal diseases, ELS, LLS and rust are economically important which are widely distributed and can cause yield losses up to 70%. Seed and soilborne diseases cause

severe seedling mortality resulting in the patchy crop, stand mostly in sandy loam soils and reduce yields from 25 to 50%. PBNB has been reported to cause yield losses up to 50% in the early infected crop. Besides reducing pod yield, it also affects the fodder quality of haulm (DGR, 2014). However, the incidence or severity of the disease may vary from season to season.

The magnitude of losses due to diseases is high all over the world. In general, the disease management programme involves the most use of pesticides and very rarely farmers go for other alternatives such as cultural, physical, biological methods etc. Hence, comprehensive pest management programme is most desirable. The management of diseases of peanut through a single biological or chemical method is very difficult and impractical. Various control measures need to be suitably integrated to cover a broad spectrum of pests. The approach, therefore, should be an integrated protection of the crop rather than integrated management of individual pest. There are a few research efforts where the pest control measures have been integrated and there appears a scope for further improvement in integrated pest management in peanut. Integration of best disease management practices for peanut diseases would be an effective approach and also the need of the hour. Appropriate use of fungicides with bio-control agents could be a cost-effective and eco-friendly approach. In the present studies, *Trichoderma harzianum* and some recommended fungicides were used for management of major soil borne and foliar fungal diseases of peanut.

Materials and Methods

The experiment was conducted in randomized block design with four replications in 15 m² (5 m x 3 m) plot size with 30 cm x 10 cm spacing at experimental farm of ICAR-

Directorate of Groundnut Research (DGR), Junagadh (N 21°31', E 70°36', 60 m asl), Gujarat (India) during the *kharif* season in 2009 and 2010 using peanut cultivar GG-20 and GG-2, respectively. The climate of the region is semi-arid with a mean annual rainfall of 850 mm. The rainfall is mainly confined during June to September. The monthly average temperature is minimum in January (12 °C) and maximum in May (42 °C). The soils of the experimental field are clayey, mixed, hyperthermic, lithic haplustepts. These soils sometimes develop shallow cracks and are very dark grey, slightly calcareous, moderately alkaline, clayey texture underlain by weathered or hard miliolitic limestone. They are also strongly alkaline, strongly stony, clayey soils.

The soils are shallow and the depth ranges from 25-50 cm. The soils are moderately well drained and are moderately permeable. They have medium runoff and are moderately eroded. The clay texture imparts good nutrient reserve capacity and good water holding capacity. Physicochemical parameters and nutrient status of experimental plots up to a depth of 0-15 cm were the organic carbon about 0.69%, EC 0.14 dSm⁻¹, pH 8.3, bulk density 1.20 g cm⁻³, water retention 28.8 at 1/3 bar and 15.4 at 15 bar, exchangeable cations (Ca 37.8, Mg 9.9, Na 1.7, K 0.3 CEC 53.2 cmol kg⁻¹ soil), ESP 3.2; CaCO₃ 3.0%, available N 72.1 kg ha⁻¹, P 17.3 kg ha⁻¹, K 176.6 kg ha⁻¹, Fe 8.74 mg kg⁻¹, Mn 9.28 mg kg⁻¹, Zn 1.5 mg kg⁻¹ and Cu 1.42 mg kg⁻¹. In spite of limited depth, soils are responsive to balanced fertilizer application particularly the inclusion of N, P, K, Zn and Fe in the fertilizer schedule. (Singh et al., 1998)

Bioagent, *T. harzianum* (NRCG, T-170) maintained at Plant Pathology section of DGR were used in the experiment. Mass multiplication of *T. harzianum* was done on the castor cake which was also a treatment.

For this *Trichoderma* culture was multiplied on sorghum grains for seven days. This was thoroughly mixed in castor cake seven days before field application. The moisture level and humidity were maintained by covering with gunny bags.

To ensure sufficient disease incidence of collar rot and stem rot, inoculums of *Aspergillus niger* @ 25 kg ha⁻¹ and *Sclerotium rolfsii* @ 100 kg ha⁻¹, multiplied on sorghum grain was added to soils before sowing and at 20 days after sowing (DAS), respectively. A spray of fungicides was scheduled as 1st spray at the first appearance of symptoms of foliar diseases followed by 2nd spray after 15 days.

Observations on foliar fungal diseases were recorded by adopting 1-9 modified scale (Subrahmanyam *et al.*, 1995) and in the case of soil borne diseases per cent incidence was computed. Per cent Soil-borne Disease incidence, Per cent efficacy of disease control (PEDC), Per cent yield increase over disease control (PIDC) was calculated by using the following formulae:

Per cent Soil-borne Disease incidence = (number of infected plant units / total number of plant units assessed) x 100

Per cent efficacy over Disease Control (PEDC) = [(Disease severity or incidence in control - Disease severity or incidence in treatment) / Disease severity or incidence in control] x 100

Per cent yield increase over Disease Control (PIDC) = [(Yield in treatments - Yield in Control) / Yield in Control] x 100

The incremental cost benefit ratio (ICBR) and Benefit: Cost Ratio (B: C ratio) of all the treatments was calculated using following formulae:

ICBR= $\frac{\text{ICBR} = \text{Additional income received (from the particular treatment)} / \text{Additional cost incurred (for the particular treatment)}}{\text{Additional cost incurred (for the particular treatment)}}$

B: C ratio= $\frac{\text{Income received (from the particular treatment)} / \text{Cost incurred (for the particular treatment)}}{\text{Cost incurred (for the particular treatment)}}$

The data were analysed following standard statistical procedures using DSAASTAT software (Onobri, 2007) and analysis of variance (ANOVA) was worked out. Data was subjected to F test and least significant difference (LSD) between means was calculated at 5% significance level ($p < 0.05$).

Results and Discussion

The results of the experiments indicated that all the treatments significantly reduced the incidence of soil-borne diseases and severity of foliar fungal diseases over the untreated control. All the treatments were found effective to maintain the initial plant population (numbers) in the field (Fig. 1). Significantly higher plant population was recorded in seed treatment with mancozeb (273) followed by seed treatment with *T. harzianum* (260) as compared to control (238). Lowest incidence of soil-borne diseases viz., aflaroot (Fig. 2), collar rot (Fig. 3) and stem rot (Fig. 4) were recorded during both the years of seed treatment with mancozeb and two spray of hexaconazole (T_2) (1.1, 8.8, and 11.6 per cent, respectively) followed by seed treatment with tebuconazole (1.5 g kg^{-1} seed) and two foliar spray of tebuconazole (T_3) (0.9, 9.0 and 11.9, respectively) in comparison to untreated control (1.7, 14.3 and 23.8, respectively) and significantly reduced the disease incidence (Table 3).

The highest PEDC (per cent efficiency over disease control) was observed in T_2 and T_3 for aflaroot (35.3 and 47.1, respectively), collar rot (38.5 and 37.1, respectively) and stem rot

(51.3 and 50.0, respectively) in comparison to other treatments applied (Table 1).

The observation on foliar diseases revealed that all the treatments had significantly reduced PDI (per cent disease incidence) in comparison to untreated control (Table 2). Seed treatment with tebuconazole and two foliar sprays of tebuconazole (T_3) was found most effective among all the treatments. The lowest PDI of ELS (Fig. 5), LLS (Fig. 6) and rust (Fig. 7) (5.9, 3.7 and 4.2, respectively) were recorded in T_3 whereas it was recorded highest (7.2, 6.8 and 6.3, respectively) in the untreated control. The PEDC (per cent efficacy over disease control) was also very high in T_3 in comparison to other treatments for ELS (17.4), LLS (45.7) and rust (33.8) (Table 1).

The data on biological yield (pod and haulm) of peanut showed the significant effect of treatments on the enhancement of the yield over the untreated control (Table 4). The highest pod yield was recorded in T_2 and T_3 (1637 kg/ha and 1627 kg/ha, respectively) whereas it was lowest in untreated control (1425 kg/ha) (Fig. 8). The haulm yield was also found highest in T_2 (3115 kg/ha) and T_3 (3097 kg/ha) in comparison to control (2743 kg/ha) (Fig. 8). Both the treatment T_2 (14.9) and T_3 (14.2) showed the highest PIDC (per cent increase over disease control) for pod yield and for haulm yield, respectively, among all the treatments (Table 1). The incremental cost benefit ratio (ICBR) calculated on the basis of prevailing market price indicated that it was highest in T_1 (1:8.3), T_2 (1:8.7) and T_3 (1:4.9). The data on ICBR showed that the cost of treatment was very important for management decision. Based on the result, it is suggested that treatment T_2 -seed treatment with mancozeb (3 g kg^{-1}) and two sprays of hexaconazole (1 ml l^{-1}) will be most economical for the management of peanut diseases.

The following treatments (fungicides) were used as seed dressing

Treatment	Treatment Detail
T ₁	Seed treatment with <i>T. harzianum</i> (10 g kg ⁻¹ seed) and two sprays of hexaconazole 5% SC (1 ml l ⁻¹)
T ₂	Seed treatment with mancozeb 75% WP (3 g kg ⁻¹) and two sprays of hexaconazole 5% SC (1 ml l ⁻¹)
T ₃	Seed treatment with tebuconazole 2 DS (1.5 g kg ⁻¹ seed) and two foliar sprays of tebuconazole 25.9% EC (1 ml l ⁻¹)
T ₄	Soil application of <i>T. harzianum</i> (4.0 kg ha ⁻¹) and castor cake (250 kg ha ⁻¹) and two sprays of hexaconazole 5% SC (1 ml l ⁻¹)
T ₅	Seed treatment with <i>T. harzianum</i> (10 g kg ⁻¹) + T ₄
T ₆	Control (without any seed treatment, only water spray)

The detail of fungicides used are given below

Common name	Trade name	Chemical name	Manufacturing company
hexaconazole/ Contaf Plus	Hexaconazole 5% SC	2-(2,4-Dichlorophenyl)-1-(1 <i>H</i> -1,2,4-triazol-1-yl)hexan-2-ol	Rallis India Limited, Mumbai
Tebuconazole	Folicur 250 EC (25.9% w/w)	(±)-1-(4-Chlorophenyl)-4,4-dimethyl-3-(1 <i>H</i> , 1,2,4-triazol-1-ylmethyl)pentan-3-ol	Bayer CropScience Ltd.
Tebuconazole	Raxil 2DS	(<i>RS</i>)- 1-(4-Chlorophenyl)- 4,4-dimethyl-3-(1 <i>H</i> , 1,2,4-triazol-1-ylmethyl)pentan- 3-ol	Bayer CropScience Ltd.
Mancozeb	TATA M-45 (mancozeb 75% WP)	zinc;manganese(2+);N-[2-(sulfidocarbothioylamino)ethyl]carbamo dithioate	Rallis India Limited, Mumbai

Table.1 Effect of different treatment on plant population and percent efficacy over disease control of major soil-borne and foliar diseases of peanut

Treatment	Initial plant population	Per cent Efficacy over Disease Control			Per cent Efficacy Over Disease Control			Per cent Yield Increase Over Disease Control	B: C Ratio	ICBR
		Aflaroot	Collar rot	Stem rot	ELS	LLS	Rust			
T ₁	260	23.5	32.9	50.4	10.5	34.0	29.9	9.6	0.68	1:8.3
T ₂	273	35.3	38.5	51.3	12.2	34.6	31.3	14.9	0.77	1:8.7
T ₃	242	47.1	37.1	50.0	17.4	45.7	33.8	14.2	0.74	1:4.9
T ₄	250	17.6	31.5	46.6	14.0	35.7	34.4	12.9	0.67	1:2.4
T ₅	246	11.8	30.1	46.2	11.1	39.5	29.1	8.1	0.62	1:1.9
T ₆	238	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.53	-
SEm±	7.36									
LSD < 0.05	21.26									

*Pooled mean of two experiments with four replication for the year 2009 and 2010

**ELS= Early leaf Spot; LLS= Late leaf spot; ICBR= Incremental cost benefit ratio

T₁: Seed treatment with *T. harzianum* (10 g kg⁻¹ seed) and two sprays of Hexaconazole (1 ml l⁻¹), T₂: Seed treatment with Mancozeb (3 g kg⁻¹) and two spray of Hexaconazole (1 ml l⁻¹), T₃: Seed treatment with Tebuconazole (1.5 g kg⁻¹ seed) and two foliar spray of Tebuconazole (1 ml l⁻¹), T₄: Soil application of *T. harzianum* (4.0 kg ha⁻¹) and castor cake (250 kg ha⁻¹) and two spray of Hexaconazole (1 ml l⁻¹), T₅: Seed treatment with *T. harzianum* (10 g kg⁻¹) + T₄; T₆: Control (without any seed treatment, only water spray)

Table.2 Analysis of variance table for effects of different treatments on severity of early leaf spot, late leaf spot, and rust (2009, 2010 and Two Year Means)

Source of variation	df ^x	Early leaf spot			Late leaf spot			Rust		
		MS ^y	F ^z	P ^z	MS ^y	F ^z	P ^z	MS ^y	F ^z	P ^z
2009										
Replication	3.0	0.4	3.9		0.3	2.8		0.6	1.3	
Treatments	5.0	0.9	8.8	<0.01	0.5	4.0	<0.05	1.1	2.1	0.1
Residual	15.0	0.1			0.1			0.5		
2010										
Replication	3.0	0.0	0.1		0.6	1.5		1.2	6.8	
Treatments	5.0	0.6	5.0	<0.01	0.7	1.7	0.2	0.5	2.7	0.1
Residual	15.0	0.1			0.4			0.2		
Two Year Means										
Replication	3.0	0.1	4.4		0.0	0.3		0.3	1.0	
Treatments	5.0	0.7	26.7	<0.01	0.6	3.8	<0.05	0.7	2.8	0.1
Residual	15.0	0.0			0.1			0.3		

^x Degree of freedom; ^y Mean square; ^z F value and probability of a greater F-value

Table.3 Analysis of variance table for effects of different treatments on incidence of aflaroot, collar rot and stem rot (2009, 2010 and Two Year Means)

Source of variation	df ^x	Aflaroot (%)			Collar rot (%)			Stem rot (%)		
		MS ^y	F ^z	P ^z	MS ^y	F ^z	P ^z	MS ^y	F ^z	P ^z
2009										
Replication	3.0	0.2	0.6		1.2	1.2		1.4	2.1	
Treatments	5.0	12.9	33.6	<0.01	12.7	12.5	<0.01	25.4	36.5	<0.01
Residual	15.0	0.4			1.0			0.7		
2010										
Replication	3.0	3.2	6.6		0.5	0.3		0.5	0.5	
Treatments	5.0	9.9	20.3	<0.01	9.4	5.6	<0.01	5.5	5.3	<0.01
Residual	15.0	0.5			1.7			1.0		
Two Year Means										
Replication	3.0	1.1	15.9		0.1	0.6		0.0	0.4	
Treatments	5.0	8.0	110.5	<0.01	2.4	16.4	<0.01	3.2	33.5	<0.01
Residual	15.0	0.1			0.1			0.1		

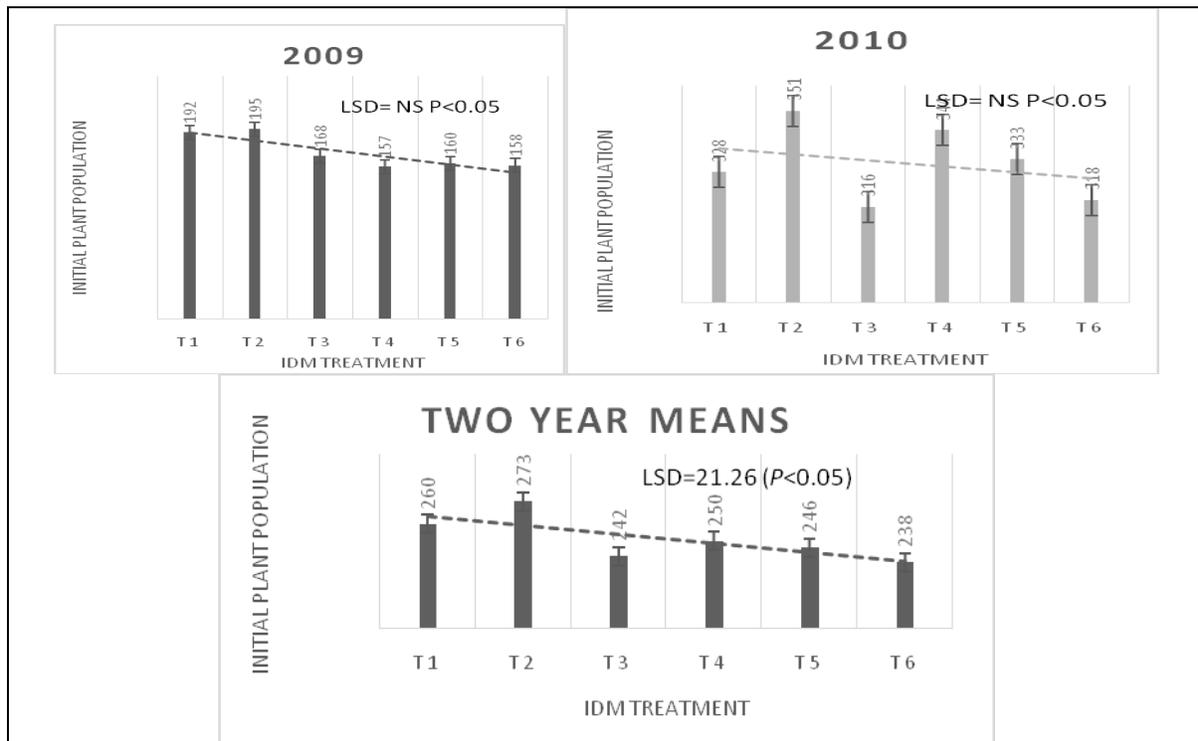
^x Degree of freedom; ^y Mean square; ^z F value and probability of a greater F-value

Table.4 Analysis of variance table for effects of different treatments on peanut yield (2009, 2010 and Two Year Means)

Source of variation	df ^x	Pod Yield (kg ha ⁻¹)			Haulm Yield (kg ha ⁻¹)		
		MS ^y	F ^z	P ^z	MS ^y	F ^z	P ^z
2009							
Replication	3.0	49289	5.3		150948	5.3	
Treatments	5.0	11463	1.2	0.3	35106	1.2	0.3
Residual	15.0	9320			28541		
2010							
Replication	3.0	48073	7.2		147224	7.2	
Treatments	5.0	56433	8.5	0.0	172827	8.5	0.0
Residual	15.0	6666			20414		
Two Year Means							
Replication	3.0	36794	6.3		112682	6.3	
Treatments	5.0	23755	4.1	0.0	72749	4.1	0.0
Residual	15.0	5802			17769		

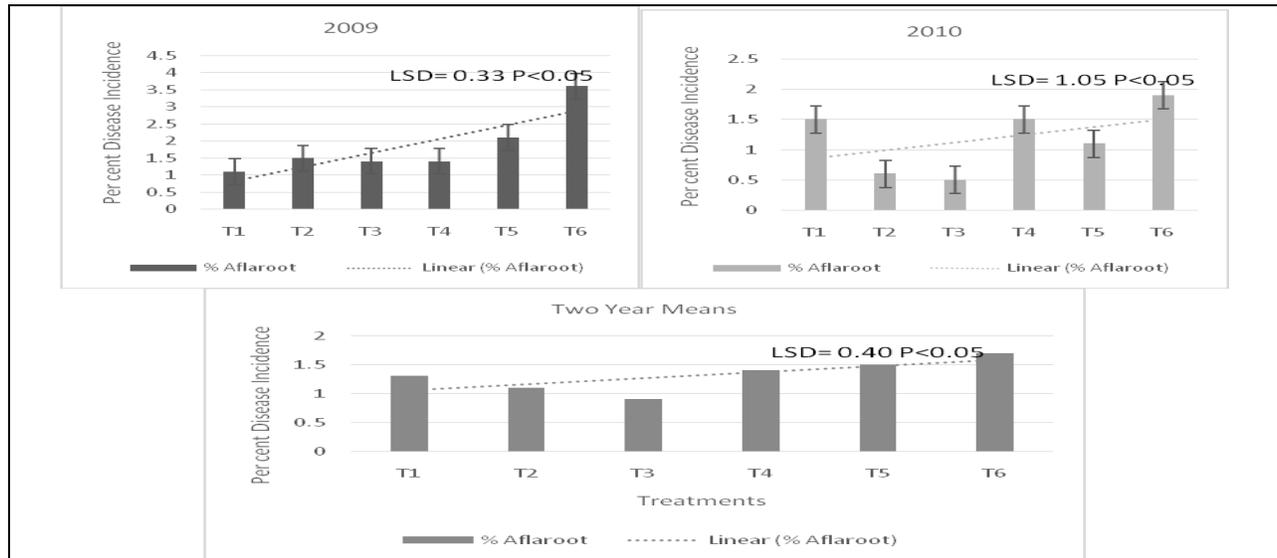
^x Degree of freedom; ^y Mean square; ^z F value and probability of a greater F-value

Fig.1 Effect of different treatments on initial plant population of Peanut. Least significant difference (LSD) values were calculated at $P < 0.05$



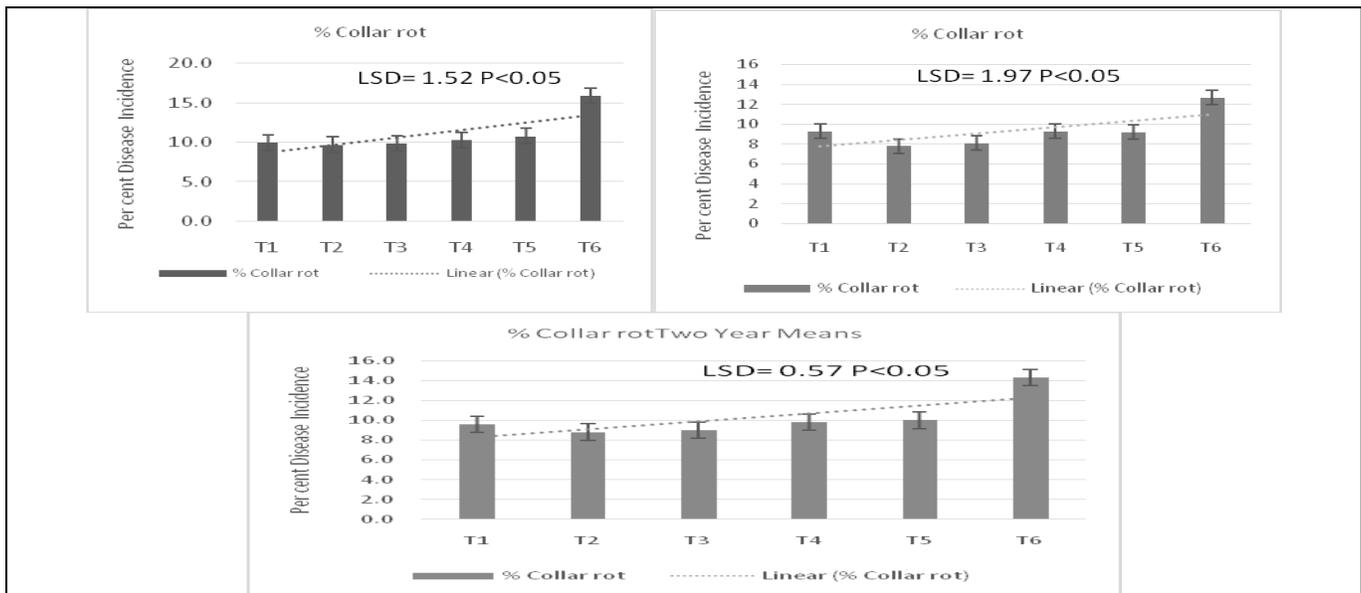
T₁: Seed treatment with *T. harzianum* (10 g kg⁻¹ seed) and two sprays of hexaconazole (1 ml l⁻¹), T₂: Seed treatment with mancozeb (3 g kg⁻¹) and two spray of hexaconazole (1 ml l⁻¹), T₃: Seed treatment with tebuconazole (1.5 g kg⁻¹ seed) and two foliar spray of tebuconazole (1 ml l⁻¹), T₄: Soil application of *T. harzianum* (4.0 kg ha⁻¹) and castor cake (250 kg ha⁻¹) and two spray of hexaconazole (1 ml l⁻¹), T₅: Seed treatment with *T. harzianum* (10 g kg⁻¹) + T₄; T₆: Control (without any seed treatment, only water spray)

Fig.2 Effect of different treatments on aflaroot incidence. Least significant difference (LSD) values were calculated at $P<0.05$



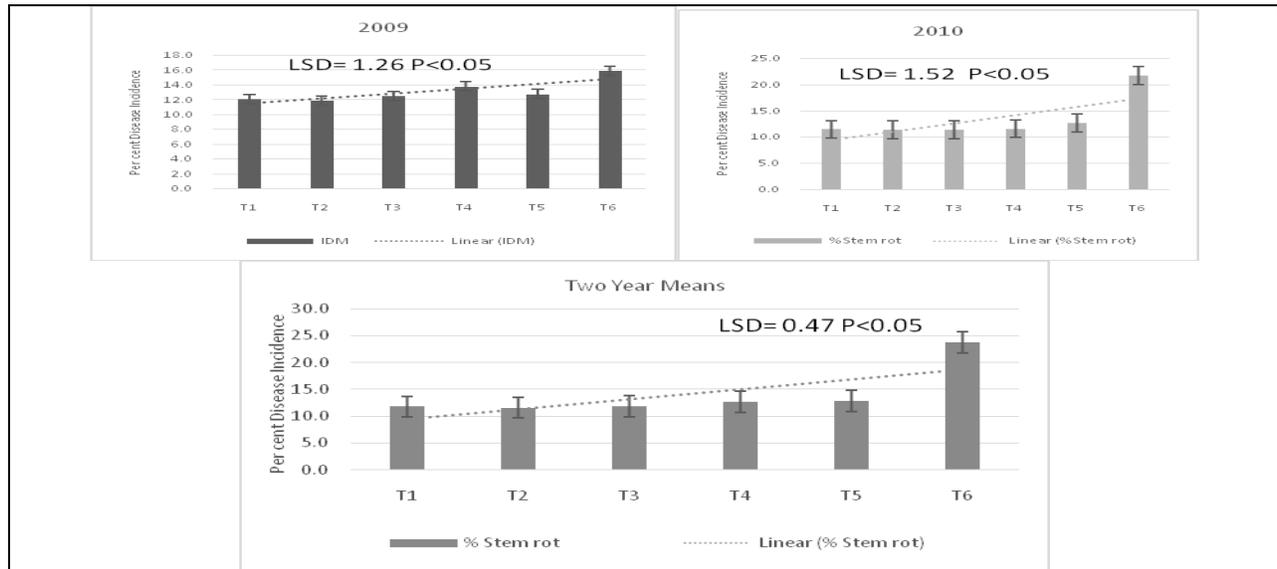
T₁: Seed treatment with *T. harzianum* (10 g kg⁻¹ seed) and two sprays of hexaconazole (1 ml l⁻¹), T₂: Seed treatment with mancozeb (3 g kg⁻¹) and two spray of hexaconazole (1 ml l⁻¹), T₃: Seed treatment with tebuconazole (1.5 g kg⁻¹ seed) and two foliar spray of tebuconazole (1 ml l⁻¹), T₄: Soil application of *T. harzianum* (4.0 kg ha⁻¹) and castor cake (250 kg ha⁻¹) and two spray of hexaconazole (1 ml l⁻¹), T₅: Seed treatment with *T. harzianum* (10 g kg⁻¹) + T₄; T₆: Control (without any seed treatment, only water spray)

Fig.3 Effect of different treatments on collar rot incidence. Least significant difference (LSD) values were calculated at $P<0.05$



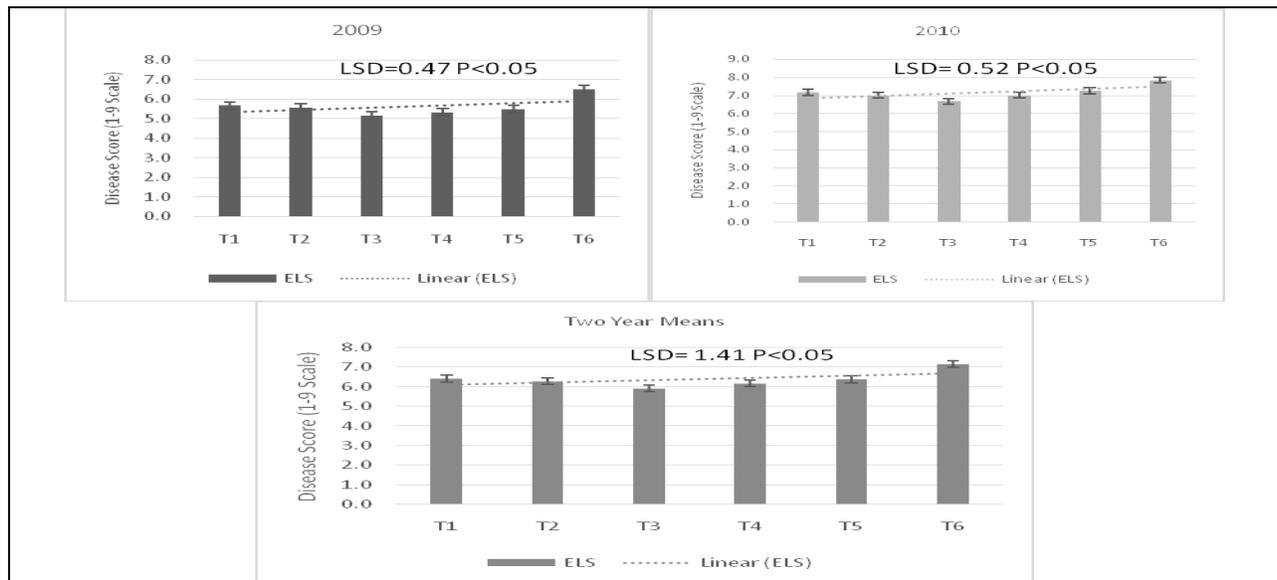
T₁: Seed treatment with *T. harzianum* (10 g kg⁻¹ seed) and two sprays of hexaconazole (1 ml l⁻¹), T₂: Seed treatment with mancozeb (3 g kg⁻¹) and two spray of hexaconazole (1 ml l⁻¹), T₃: Seed treatment with tebuconazole (1.5 g kg⁻¹ seed) and two foliar spray of tebuconazole (1 ml l⁻¹), T₄: Soil application of *T. harzianum* (4.0 kg ha⁻¹) and castor cake (250 kg ha⁻¹) and two spray of hexaconazole (1 ml l⁻¹), T₅: Seed treatment with *T. harzianum* (10 g kg⁻¹) + T₄; T₆: Control (without any seed treatment, only water spray)

Fig.4 Effect of different treatments on stem rot incidence. Least significant difference (LSD) values were calculated at $P<0.05$



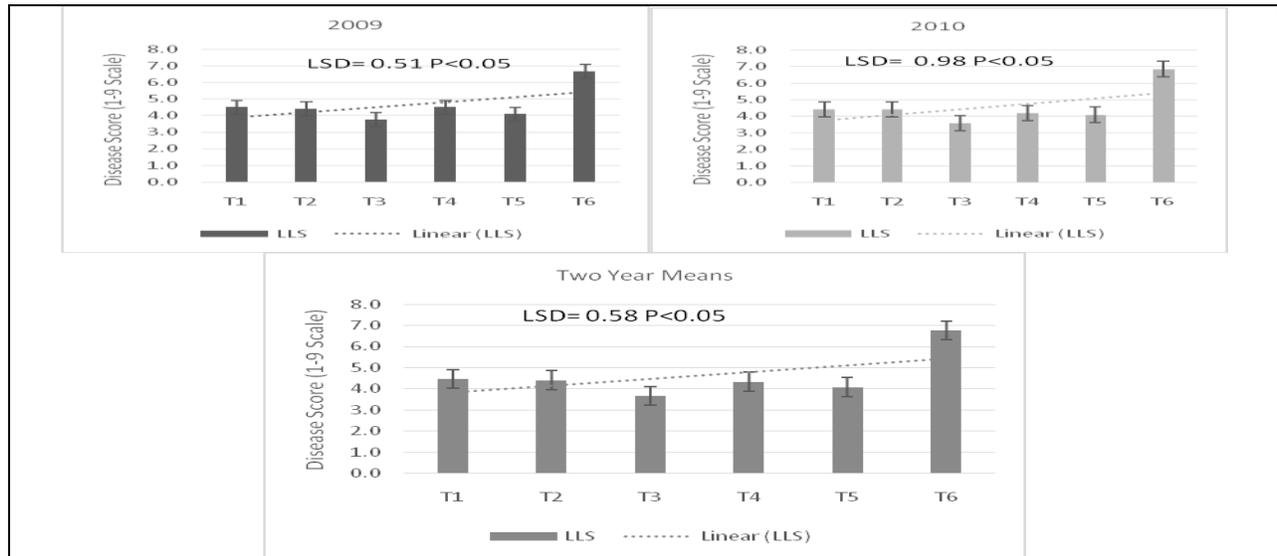
T₁: Seed treatment with *T. harzianum* (10 g kg⁻¹ seed) and two sprays of hexaconazole (1 ml l⁻¹), T₂: Seed treatment with mancozeb (3 g kg⁻¹) and two spray of hexaconazole (1 ml l⁻¹), T₃: Seed treatment with tebuconazole (1.5 g kg⁻¹ seed) and two foliar spray of tebuconazole (1 ml l⁻¹), T₄: Soil application of *T. harzianum* (4.0 kg ha⁻¹) and castor cake (250 kg ha⁻¹) and two spray of hexaconazole (1 ml l⁻¹), T₅: Seed treatment with *T. harzianum* (10 g kg⁻¹) + T₄; T₆: Control (without any seed treatment, only water spray)

Fig.5 Effect of different treatments on early leaf spot (ELS) severity. Least significant difference (LSD) values were calculated at $P<0.05$



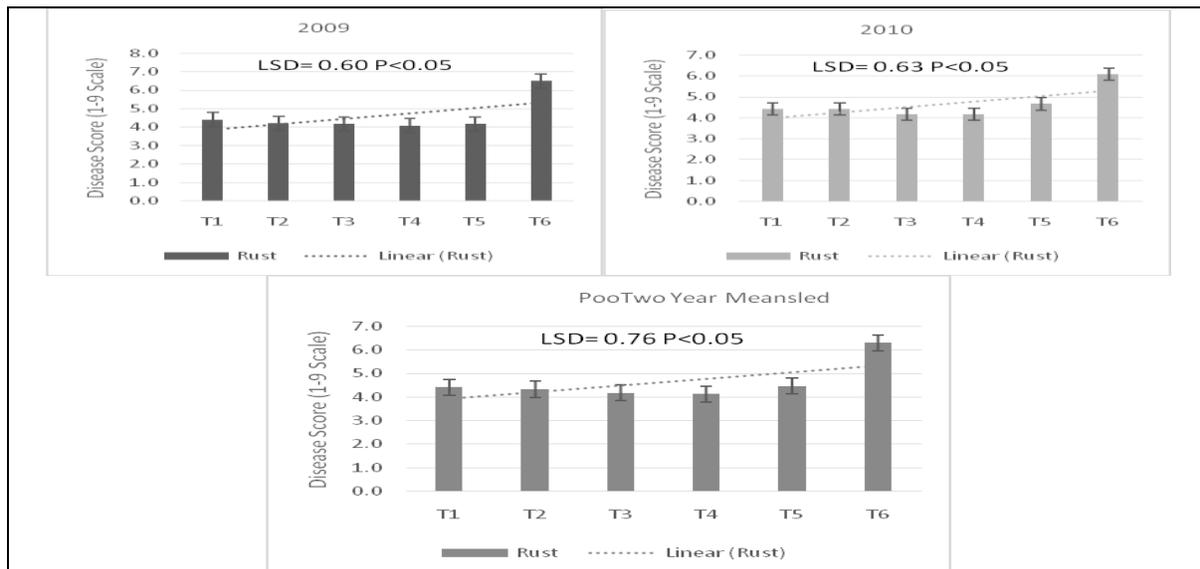
T₁: Seed treatment with *T. harzianum* (10 g kg⁻¹ seed) and two sprays of hexaconazole (1 ml l⁻¹), T₂: Seed treatment with mancozeb (3 g kg⁻¹) and two spray of hexaconazole (1 ml l⁻¹), T₃: Seed treatment with tebuconazole (1.5 g kg⁻¹ seed) and two foliar spray of tebuconazole (1 ml l⁻¹), T₄: Soil application of *T. harzianum* (4.0 kg ha⁻¹) and castor cake (250 kg ha⁻¹) and two spray of hexaconazole (1 ml l⁻¹), T₅: Seed treatment with *T. harzianum* (10 g kg⁻¹) + T₄; T₆: Control (without any seed treatment, only water spray)

Fig.6 Effect of different treatments on late leaf spot (LLS) severity. Least significant difference (LSD) values were calculated at $P<0.05$



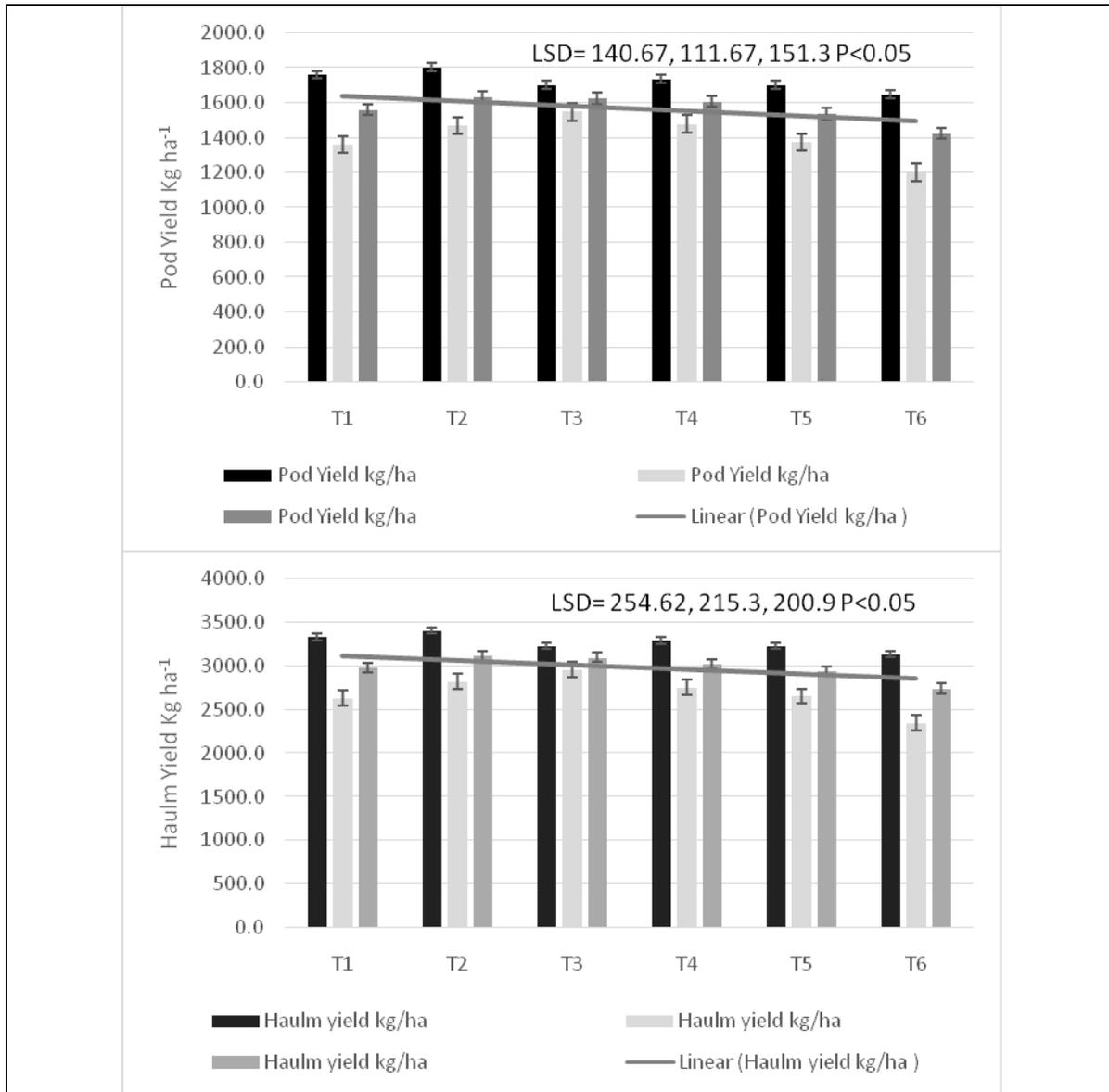
T₁: Seed treatment with *T. harzianum* (10 g kg⁻¹ seed) and two sprays of hexaconazole (1 ml l⁻¹), T₂: Seed treatment with mancozeb (3 g kg⁻¹) and two spray of hexaconazole (1 ml l⁻¹), T₃: Seed treatment with tebuconazole (1.5 g kg⁻¹ seed) and two foliar spray of tebuconazole (1 ml l⁻¹), T₄: Soil application of *T. harzianum* (4.0 kg ha⁻¹) and castor cake (250 kg ha⁻¹) and two spray of hexaconazole (1 ml l⁻¹), T₅: Seed treatment with *T. harzianum* (10 g kg⁻¹) + T₄; T₆: Control (without any seed treatment, only water spray)

Fig.7 Effect of different treatments on rust severity. Least significant difference (LSD) values were calculated at $P<0.05$



T₁: Seed treatment with *T. harzianum* (10 g kg⁻¹ seed) and two sprays of hexaconazole (1 ml l⁻¹), T₂: Seed treatment with mancozeb (3 g kg⁻¹) and two spray of hexaconazole (1 ml l⁻¹), T₃: Seed treatment with tebuconazole (1.5 g kg⁻¹ seed) and two foliar spray of tebuconazole (1 ml l⁻¹), T₄: Soil application of *T. harzianum* (4.0 kg ha⁻¹) and castor cake (250 kg ha⁻¹) and two spray of hexaconazole (1 ml l⁻¹), T₅: Seed treatment with *T. harzianum* (10 g kg⁻¹) + T₄; T₆: Control (without any seed treatment, only water spray)

Fig.8 Effect of different treatments on pod yield (kg ha^{-1}) and haulm yield (kg ha^{-1}) of Peanut. Least significant difference (LSD) values were calculated at $P < 0.05$



T₁: Seed treatment with *T. harzianum* (10 g kg^{-1} seed) and two sprays of hexaconazole (1 ml l^{-1}), T₂: Seed treatment with mancozeb (3 g kg^{-1}) and two spray of hexaconazole (1 ml l^{-1}), T₃: Seed treatment with tebuconazole (1.5 g kg^{-1} seed) and two foliar spray of tebuconazole (1 ml l^{-1}), T₄: Soil application of *T. harzianum* (4.0 kg ha^{-1}) and castor cake (250 kg ha^{-1}) and two spray of hexaconazole (1 ml l^{-1}), T₅: Seed treatment with *T. harzianum* (10 g kg^{-1}) + T₄; T₆: Control (without any seed treatment, only water spray)

The results of the experiments indicated that all the treatments significantly reduced the incidence of soil-borne diseases and severity of foliar fungal diseases over the untreated control. Dandnaik *et al.*, (2009) reported that hexaconazole as seed treatment showed the

highest seed germination (75%). Sharma and Gour (2009) noted that seed treatment with carbendazim and propiconazole also resulted in increased germination and seedling vigour of pea. Significantly higher germination percent (93) was observed with carbendazim

as a seed treatment against *Macrophomina phaseolina* in cluster bean. Shirao *et al.*, (2009) reported that seed treatment with thiram + carbendazim + *Trichoderma* sp. effectively reduced the mortality (75%) due to root and collar rot of soybean as compared to control. Similarly, seed treatments with carbendazim, captan, and carbendazim + captan were found effective and at par in recording significantly higher pre- and post-emergence mortality due to collar rot in soybean (Pawar *et al.*, 2009). Sreedevi *et al.*, (2011) reported that the growth of peanut plants with the antagonist alone or in combination with pathogen was greater than in plants inoculated with pathogen alone. Sai *et al.*, (2010), reported *Trichoderma* isolates TG2 was significantly superior over others in inhibiting the mycelial growth of *S. rolfisii* to the extent of 67.83%. Harsukh *et al.*, (2011) also reported that seed treatment with *Trichoderma* reduced the disease incidence in susceptible and tolerant varieties at 15 DAS, under *A. niger* infection.

In the present studies, the lowest incidence of soil-borne diseases was recorded during both the years of seed treatment with mancozeb and two sprays of hexaconazole and seed treatment with tebuconazole and two foliar sprays of tebuconazole in comparison to untreated control and significantly reduced the disease incidence. Johnson and Subramanyam (2010) and Dandnaik *et al.*, (2009) also reported that seed treatment with hexaconazole showed the maximum reduction of stem rot incidence (24%) in peanut. Minimum incidence of pre-emergence rot and post-emergence seedling rot (13%) were recorded in treatment with carbendazim and thiophanete methyl compared to control (43 and 49 percent, respectively) in cluster bean (Jaiman *et al.*, 2009). Gour and Sharma (2010) also reported that sprays of Folicur 250 EW in the field, along with two standard controls, i.e. Tilt 25 EC and contaf 5 EC

showed notable efficacy, resulting in low PDI of 8.7 and 16.4% and PEDC of 87.9 and 77.3%, respectively, after 30 days. Tilt 25 EC and Contaf 5 EC also showed disease control efficacy of 68.9 and 65.6%, respectively. In the uninoculated control plots, the disease severity (PDI) was 28.5%. All the concentrations of Folicur 250 EW were significantly superior in PEDC to the standard controls, Tilt 25 EC (68.9%) and contaf 5 EC (65.6%). Augusto and Brenneman (2012) studied the movement of systemic fungicides in three terminal, fully expanded leaves of primary lateral branches of 'Tifrunner' peanut treated with prothioconazole + tebuconazole (Provost, 0.29 kg a.i./ha), azoxystrobin (Abound, 0.31 kg a.i./ha), or flutolanil (moncut, 0.79 kg a.i./ha) in field experiments. Their results demonstrated acropetal protection by all the fungicides evaluated and indicated that prothioconazole + tebuconazole or prothioconazole applied to foliage can sometimes reduce diseases in the lower, non-treated portions of the plant.

The highest PEDC (per cent efficiency over disease control) was observed in T₂ and T₃ for soil borne diseases in comparison to other treatments applied. These results were also in concurrence with Devi and Prasad (2009) who reported that combined effect of seed treatment with *T. viride* and captan resulted in significant reduction of collar rot and combination of antagonist and fungicide also improved the growth parameters like the length of the plant, biomass and yield besides decreasing the disease incidence. Muhammad and Bdliya (2011) evaluated the effects of variety and fungicide (mancozeb) on *Cercospora* leaf spot disease of peanut and found that disease incidence and disease severity were significantly higher in 'Ex-Dakar' variety and untreated seeds, thus had lower yields. Higher seed and haulm yield were recorded by RRB, ICGV-86024, as well as 2 and 4 kg ha⁻¹ as a result of lower disease

incidence and severity. Wann *et al.*, (2011) also evaluated the efficacy of three approved fungicides for leaf spot control on peanut under organic management. Copper sulfate+ *Bacillus subtilis* reduced leaf spot defoliation compared to the control. Yet, all three fungicides improved yields under heavy leaf spot pressure. Combining high-yielding, disease-resistant cultivars and organically approved fungicides can significantly reduce leaf spot and improve yield potential of peanut under organic management. Augusto and Brenneman (2011) reported that night and day applications of a systemic fungicide for leaf spot disease control were similar across post-spray irrigation, but pyraclostrobin and prothioconazole + tebuconazole had the lowest disease score. Interaction of fungicide, application timing, and post-spray irrigation was significant for stem rot and yield. Night application of prothioconazole + tebuconazole, flutolanil + propiconazole or pyraclostrobin showed the most increase in stem rot control and yield compared with day application among the evaluated fungicides, but the positive effects on stem rot control and yield were minimal with post-spray irrigation.

The observation on foliar diseases revealed that all the treatments had significantly reduced PDI (per cent disease incidence) in comparison to untreated control. Seed treatment with tebuconazole and two foliar sprays of tebuconazole was found most effective among all the treatments. The lowest PDI of foliar diseases were recorded in T₃ whereas it was recorded highest in the untreated control. The PEDC (per cent efficacy over disease control) was also very high in T₃ in comparison to other treatments. Hagan *et al.*, (2010) also reported azoxystrobin, chlorothalonil, and tebuconazole, for the control of ELS and stem rot and found the incidence of these diseases were lower with the application of

azoxystrobin than chlorothalonil in the 2 out of 3 year's programs. The azoxystrobin program significantly increased yield compared with the chlorothalonil or tebuconazole. The yield was also higher in the tebuconazole treatment compared with chlorothalonil. Grichar *et al.*, (2010) conducted a field experiment to determine both foliar and soil-borne disease control and peanut response to prothioconazole + tebuconazole (Provost) fungicide treatments when compared with other fungicides commonly used in peanut in different peanut growing areas of Texas. ELS control with prothioconazole + tebuconazole was comparable with azoxystrobin (Abound), chlorothalonil (Bravo), or tebuconazole (Folicur) and incidence of the disease was reduced at least 36% relative to the untreated check. Southern stem rot control with prothioconazole + tebuconazole was comparable to azoxystrobin or pyraclostrobin (Headline) and all fungicides reduced disease incidence at least 60% when compared to the untreated check. Peanut yields with prothioconazole + tebuconazole were increased at least 85% over the untreated check when ELS was present and at least 34% over those of the untreated check when Southern stem rot was present.

The data on biological yield (pod and haulm) of peanut showed the significant effect of treatments on the enhancement of the yield over the untreated control. The highest pod yield was recorded in T₂ and T₃ whereas it was lowest in the untreated control. The haulm yield was also found highest in T₂ and T₃ in comparison to control. Both the treatment T₂ and T₃ showed the highest PIDC (per cent increase over disease control) for pod yield and for haulm yield, respectively, among all the treatments. The results confirmed the findings of Johnson and Subramanyam (2010) and Dandnaik *et al.*, (2009) who reported that seed treatment with

hexaconazole gave higher pod yield of 1400 kg/ha in peanut. The effect of seed treatment in increasing the yields was reported by several workers viz., De *et al.*, (2003) with carbendazim + thiram against wilt of linseed, Ramkishore and Singh (2008) with carbendazim against wilt of linseed, Jaiman and Jain (2008) with carbendazim against blight and root rot of cluster bean, Dudi and Lodha (2003) with carbendazim against seedling diseases of peanut and Dutta and Das (2002) with mancozeb against collar rot of tomato. Gour and Sharma (2010) also reported that for pod yield, there was no significant difference in the two concentrations of Folicur 250 EW (2550 and 2510 kg/ha at 187.50 and 156.25 g a.i./ ha, respectively compared to 1720 kg/ha in the inoculated control). The highest percentage increase of pod (48.2%) was observed in Folicur 250 EW at 187.5 g a.i./ha, followed by at 156.25 g a.i./ha (45.9%). In applications of Tilt 25 EC and Contaf 5 EC, the percentage increase in pod yield was 36.0 and 35.5%, respectively compared to the inoculated control. The data showed that the cost of treatment are very important to manage the disease so it is suggested that treatment T₂-seed treatment with mancozeb (3 g kg⁻¹) and two sprays of hexaconazole (1 ml l⁻¹) will be most suitable for management of peanut disease.

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